

The Effect of Caffeic Acid Phenethyl Ester (CAPE) Against Cholestatic Liver Injury in Rats

Sacid Coban, M.D.,^{*,1} Fahrettin Yildiz, M.D.,[‡] Alpaslan Terzi, M.D.,[‡] Behcet Al, M.D.,[†]
Dincer Ozgor, M.D.,[§] Cengiz Ara, M.D.,[§] Alaattin Polat, M.D.,^{||} and Mukaddes Esrefoglu, M.D.[¶]

^{*}Department of General Surgery; [†]Department of Emergency Medicine, Gaziantep University School of Medicine, Gaziantep, Turkey;

[‡]Department of General Surgery, Harran University School of Medicine, Sanliurfa, Turkey; [§]Department of General Surgery;

^{||}Department of Physiology; and [¶]Department of Histology and Embryology, Inonu University School of Medicine, Malatya, Turkey

Submitted for publication July 5, 2008

Objectives. Caffeic acid phenethyl ester (CAPE) has been subjected to considerable investigations that have revealed its antioxidant and anti-inflammatory activities in different conditions. But there is not a previous investigation about its effect on cholestatic liver injury. The aim of this study was to investigate the effect of CAPE in rat liver against cholestatic liver injury induced by bile duct ligation.

Methods. Swiss-albino rats were recruited in the study as follows; Group 1 rats subjected to simple laparotomy known as the sham group; Group 2 rats subjected to bile duct ligation (BDL); Group 3 bile duct ligated rats treated with CAPE. The third group received CAPE (10 $\mu\text{mol/kg}$) intraperitoneally daily throughout 14 d.

Results. Data showed a decrease in γ glutamyl transferase (GGT), aspartate aminotransferase (AST), and alanine aminotransferase levels (ALT) of the CAPE treated rats, compared with BDL group ($P < 0.001$, $P < 0.01$, and $P < 0.02$, respectively). In the CAPE treated rats, tissue levels of malondialdehyde (MDA) and myeloperoxidase (MPO) were significantly lower than that of the BDL group ($P < 0.001$). The levels of glutathione (GSH) in CAPE treated rats were significantly higher than that of BDL group ($P < 0.001$). In CAPE treated group, the levels of interleukin-1 α (IL-1 α) and interleukin-6 (IL-6) were significantly lower than that of BDL group ($P < 0.03$, $P < 0.02$, respectively). Administration of CAPE in the rats with biliary obstruction resulted in inhibition of necro-inflammation.

Conclusion. These results suggest that treatment of CAPE maintains antioxidant defenses, reduces oxidative liver injury, cytokine damage, and necro-inflammation in bile duct ligated rats. Thus, CAPE seems to be a promising agent for the attenuation of cholestatic liver injury. © 2010 Elsevier Inc. All rights reserved.

Key Words: caffeic acid phenethyl ester (CAPE); bile duct ligation; cholestatic liver injury.

INTRODUCTION

The pathogenesis of liver injury induced by bile duct ligation (BDL) is poorly understood. There are many experimental and clinical data that show the important role of reactive oxygen species (ROS) in the pathogenesis of hepatic damage and cholestasis produced by biliary obstruction [1–3]. The bile acid concentrations increase in rats after BDL and induce lipid peroxidation, which is probably related to the stimulation of phagocytic activity in polymorphonuclear phagocytes and inflammatory cells [4]. A recent study showed that hepatic neutrophil infiltration is closely related to an increase in hepatic lipid peroxidation in bile duct ligated rats [5].

CAPE is an active component of honeybee propolis extracts and has been used for many years as a folk medicine. It has anti-inflammatory, immunomodulatory, antiproliferative, and antioxidant properties and have been shown to inhibit lipo-oxygenase activities as well as to suppress lipid peroxidation [6–9]. Also, it has been previously shown that intraperitoneal administration of caffeic acid phenethyl ester in bile duct-ligated rats

¹ To whom correspondence and reprint requests should be addressed at Department of General Surgery, Gaziantep University, Medical Faculty, 27310, Gaziantep, Turkey. E-mail: sacitcoban@yahoo.com.

reduces intestinal oxidative [10]. To date, there is no reported study regarding the effect of CAPE on cholestatic liver damage. The aim of this study was to evaluate whether CAPE administration would protect against cholestatic liver injury in rats with BDL. To assess the effect of CAPE in bile duct ligated rats, we measured the activities of glutathione (GSH), malondialdehyde (MDA), myeloperoxidase (MPO), interleukin-1alpha (IL-1 α), interleukin-6 (IL-6), and tumor necrosis factor-alpha (TNF- α). Also we examined histopathological changes in liver.

MATERIALS AND METHODS

Experimental Conditions

Three-month-old male Swiss-albino rats, weighing 300 to 350 g, were included in the study. The study was approved by Inonu University Ethics Committee.

Rats were kept in stainless steel cages, allowed access to food and water *ad libitum*, and quarantined 7 d before surgery. Food was withheld 8 h prior to surgery, but free access to water was allowed. Rats were subjected to a controlled environment regarding temperature and humidity and a 12-h light-dark cycle. All surgical procedures were performed while the rats were under intraperitoneal ketamine (50 mg/kg) and xylazine HCl (10 mg/kg) anesthesia.

A total of 21 rats were divided into three groups: Group 1, BDL ($n = 7$); Group 2, BDL plus CAPE ($n = 7$), and Group 3, sham-operated group ($n = 7$), subjected to simple laparotomy. Acute cholestatic liver injury was induced in animals by double ligation and division of the bile duct [11]. Abdominal layers were closed with appropriate suture materials. All animals were maintained under the same conditions after surgery. The CAPE was synthesized by the standard method of Grunberger [12], and administered intraperitoneally once a day at a dose of 10 $\mu\text{mol/kg}$, from 25 $\mu\text{mol/mL}$ solution in 1% ethanol.

Animals in CAPE group were treated with CAPE (10 $\mu\text{mol/kg/d}$, i.p) throughout 14 d. To eliminate complications arising from the diurnal effects, all rats were sacrificed under anesthesia at the same time of the day and liver was rapidly removed. Blood samples were taken by vena cava inferior puncture. A part of the liver was preserved in formalin for histological examination and the remainder was stored at -30°C for the analyses of MDA, MPO, GSH, and cytokines (IL-1 α , IL-6, TNF- α) levels.

Biochemical Analyses

The plasma was used to measure total bilirubin to reflect mechanical obstruction, AST and ALT to reflect the degree of hepatocellular injury, and ALP and GGT to reflect the degree of cholestasis. The plasma activities of total bilirubin, AST, ALT, ALP, and GGT were estimated by commercially available kits (Olympus Diagnostica GmbH, Dublin, Ireland). The liver tissues were homogenized and the MDA contents of homogenates were determined spectrophotometrically [13].

Myeloperoxidase activity was determined using a 4- aminoantipyrine/phenol solution as the substrate for MPO-mediated oxidation by H_2O_2 , and changes in absorbance at 510 nm (A_{510}) were recorded [14]. One unit of MPO activity was defined as the amount of protein that degrades 1 $\mu\text{mol H}_2\text{O}_2 \text{ min}^{-1}$ at 25 $^\circ\text{C}$. Results were presented as mUg^{-1} protein.

Results were expressed as umol/g tissue. GSH was determined by the spectrophotometric method, which was based on the use of Elman's reagent [15]. Results were expressed as nmol/g tissue.

Tissue homogenates were prepared using a homogenizer IKA Ultra-Turnax ($2 \times 45 \text{ s}$, 0°C) in 0.5 m TRIS/1.5m NaCl/50mM $\text{CaCl}_2/2 \text{ mM}$ sodium azide buffer at $\text{pH} = 7$. The homogenates were

then centrifuged at 15,000 g for 15 min at a temperature $+4^\circ\text{C}$ and the supernatants were used for ELISA. Rat IL-1 α , Rat IL-6, and Rat TNF- α (Biosource Immunoassay Kit, Camarillo, CA) levels were measured using a Sandwich enzyme-linked immunosorbent assay (ELISA) protocol supplied by the manufacturer of the antibodies and resultant optical density determined using a microplate reader at 450 nm. Results were expressed as pg/g tissue.

Histopathologic Evaluation

After the macroscopic findings were noted, the livers were promptly removed and processed for histologic and biochemical examination. The right lobe of the liver was divided into two pieces. The first samples were placed in 10% formalin for histopathologic examination by light microscopy. Liver tissues were fixed in 10% formalin and embedded in paraffin. For histopathologic evaluation, 4 mm slides were stained with hematoxylin-eosin. Sections were scored by an independent observer blinded to the experimental protocol. The following lesions were scored according to the modified histological activity index (HAI) [16, 17]: portal inflammation, focal necrosis, confluent necrosis, piecemeal necrosis, apoptosis, and focal inflammation.

Statistical Analysis

The results were statistically analyzed by the Kruskal-Wallis H test. The differences between groups were evaluated by the Mann-Whitney U test followed by *t*-test with Bonferroni correction when indicated. The results were given in the text as mean \pm STD for all comparisons; statistical significance was defined as $P < 0.05$.

RESULTS

All of the animals survived until the end of the experiment. Jaundice was observed in the visceral and parietal peritoneum of all animals except those of the sham group. The livers were enlarged and the bile ducts above the obstruction point were dilated. Animals of the sham group did not show any alteration after surgery.

The results of MDA, GSH, and MPO are shown in Table 1. In the CAPE treated rats, the MDA and MPO levels were significantly lower, and the levels of GSH were significantly higher than that of the BDL group ($P < 0.001$ and $P < 0.001$, respectively). Total bilirubin,

TABLE 1
Tissue Levels of MDA, GSH, and MPO Activities in the Groups

Groups	MDA (nmol/g tissue)	GSH (nmol/g tissue)	MPO (mUg^{-1} protein)
I. Sham ($n = 7$)	68.35 \pm 12.9	1.37 \pm 0.8	6.21 \pm 1.1
II. BDL ($n = 7$)	155.42 \pm 9.58	1.31 \pm 0.1	17.29 \pm 2.3
III. BDL + CAPE ($n = 7$)	119.38 \pm 16.7	2.75 \pm 0.5	8.79.1 \pm 1.4
<i>P</i> values*			
I versus II	0.001	0.53	0.001
II versus III	0.001	0.001	0.001

BDL=bile duct ligation; CAPE=caffeic acid phenethyl ester; MDA=malondialdehyde; GSH=reduced glutathione; MPO=myeloperoxidase.

* $P < 0.05$ was considered to be statistically significant.

TABLE 2
Plasma Total Bilirubin, AST, ALT, AP, and GGT Levels in the Groups

Groups	Total bilirubin (mg/dL)	AST (U/L)	ALT (U/L)	ALP (U/L)	GGT (U/L)
Sham (<i>n</i> = 7)	0.18 ± 0.17	122.8 ± 13.9	54.4 ± 7.5	353.5 ± 27.9	2.5 ± 0.9
BDL (<i>n</i> = 7)	9.5 ± 3.5	498.00 ± 121.4	157.5 ± 22.2	1663.2 ± 401.3	12.1 ± 2.7
BDL+CAPE (<i>n</i> = 7)	6.8 ± 1.6	315.7 ± 96.6	98.7 ± 45.7	1564.2 ± 913.4	4.7 ± 2.8
<i>P</i> values*					
I versus II	0.001	0.001	0.001	0.01	0.02
II versus III	0.12	0.01	0.02	0.31	0.001

BDL=bile duct ligation; CAPE=caffeic acid phenethyl ester; AST=aspartate aminotransferase; ALT=alanine aminotransferase; ALP=alkaline phosphatase; GGT=Gama glutamyl transferase.

**P* < 0.05 was considered to be statistically significant.

AST, ALT, ALP, and GGT levels in rats with BDL were significantly higher than that of sham group (Table 2). In the CAPE treated rats, the levels of AST, ALT, and GGT were significantly lower than that of the BDL group (*P* < 0.01, *P* < 0.02, and *P* < 0.001, respectively). Decreases of total bilirubin and ALP levels was not statistically significant in CAPE treated group compared with BDL group (*P* = 0.12, *P* = 0.31, respectively). The results of IL-1 α , IL-6, and TNF- α are shown in Table 3. In the CAPE treated rats, the liver levels of IL-1 α , IL-6, and were found to be significantly lower when compared to the BDL group (*P* < 0.03, *P* < 0.02, respectively). Although levels of TNF- α were decreased in CAPE injected rats compared to BDL group, the difference was not statistically significant (*P* = 0.053).

Microscopic Findings

Animals of the sham group did not present any histological alterations. The liver specimens of the BDL group showed prominent lobular and portal changes. The degree of necro-inflammation (focal necrosis, confluent necrosis, piecemeal necrosis, focal and portal inflammation, and apoptosis) showed significant difference between the BDL and BDL+CAPE groups (*P* < 0.05). HAI scores of the groups are summarized in Table 4. In the BDL group, the histopathological changes

including necrosis (Fig. 1), bile duct proliferation, fibrosis, periportal and parenchymal polymorphonuclear leucocyte (PNL), and lymphocyte infiltration were prominent (Figs. 2 and 3). There were numerous large areas of coagulation necrosis, randomly distributed (Fig.1). Fibrotic reaction was usually accompanied with inflammatory cells in the periphery of the majority of the portal space areas. In some of the portal areas, cholangitis was observed. Bile ducts were filled with pus cells, which also infiltrated their walls (Fig. 3). In the BDL+CAPE group, histopathological evidence of parenchymatous injury was markedly reduced (Figs. 4 and 5). Relatively small, scattered necrotic hepatocyte groups between normal-appearing parenchymal cells were observed (Fig. 4). Portal fibrosis was not evident. Bile duct proliferation was still prominent (Fig. 5).

DISCUSSION

The results of the present study demonstrate that treatment of CAPE markedly ameliorates the liver injury in rats subjected to BDL. Several reports showed that oxidative stress associated with lipid peroxidation is involved in the development of cholestatic liver injury in bile duct ligated rats [3, 18, 19]. The present study indicates that intraperitoneal administration of CAPE at

TABLE 3
Tissue Levels of TNF- α , IL-6, and IL-1 α Activities in the Groups

Groups	TNF- α (pg/g tissue)	IL-6 (pg/g tissue)	IL-1 α (pg/g tissue)
I. Sham (<i>n</i> = 7)	15314.00 ± 4245.57	4941.14 ± 1111.90	1834.28 ± 578.57
II. BDL (<i>n</i> = 7)	35112.85 ± 17142.95	11678.71 ± 6477.56	4493.85 ± 2158.76
III. BDL+CAPE (<i>n</i> = 7)	19284.00 ± 5540.03	6104.57 ± 1632.42	2609.42 ± 780.41
<i>P</i> values*			
I versus II	0.01	0.01	0.004
II versus III	0.053	0.02	0.03

BDL=bile duct ligation; CAPE=caffeic acid phenethyl ester; IL-1 α =interleukin-1 alpha; IL-6=interleukin-6; TNF- α =tumor necrosis factor-alpha.

**P* < 0.05 was considered to be statistically significant.

TABLE 4

Scores for Necro-Inflammation (Focal Necrosis, Confluent Necrosis, Piecemeal Necrosis, Focal and Portal Inflammation, and Apoptosis) of Groups Using Modified HIA Grading System (Mean \pm SE)

Groups	Scores for necro-inflammation
Sham ($n = 7$)	0.285 \pm 0.184
BDL ($n = 7$)	8.714 \pm 1.523*
BDL+CAPE ($n = 7$)	5.285 \pm 0.860**

BDL=bile duct ligation; CAPE=caffeic acid phenethyl ester; HAI=histological activity index.

* $P < 0.005$ versus sham.

** $P < 0.05$ versus BDL.

a dose of 10 $\mu\text{mol/kg/d}$ reduced the already increased levels of plasma AST, ALT, and GGT after bile duct ligation. Additionally, CAPE decreased oxidative stress and the increased levels of IL-1 α , IL-6, and inhibited the occurrence of necro-inflammation (focal necrosis, confluent necrosis, piecemeal necrosis, focal and portal inflammation, and apoptosis), seen in bile duct ligated rats.

In the current study, an increase in neutrophil infiltration in liver tissues, which was assessed by measuring hepatic activity, was established [20]. Neutrophil infiltration and activation are involved in the development of cholestatic liver injury in bile duct ligated rats [21, 22]. Our results are in agreement with previous works reporting high levels of MPO after the ligation of bile duct [20, 23, 24]. In the present study, levels of MPO in the CAPE treated rats were significantly lower than that of the BDL group. Reductions in MPO levels in the CAPE treated rats may probably be due to its antioxidant, free-radical-scavenging effect [8, 9], and inhibition of neutrophil infiltration [25]. This finding shows that CAPE exerts a preventive effect on

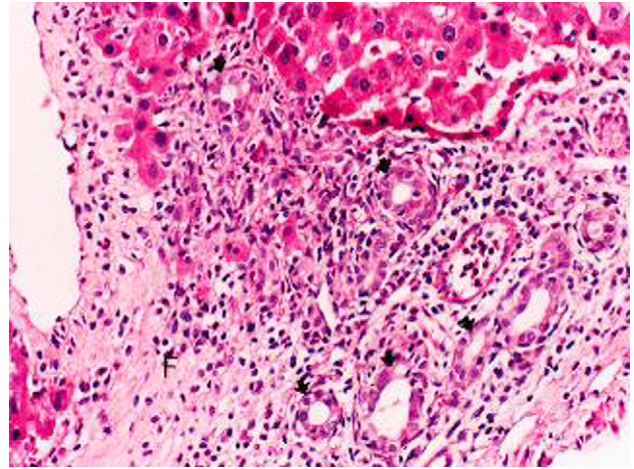


FIG. 2. BDL group. Portal fibrosis (F) with bile duct proliferation (arrows) and cell infiltration are observed. H and E; $\times 20$. (Color version of figure is available online.)

cholestatic liver injury by inhibiting enhanced neutrophil infiltration into the liver tissue.

MDA is a secondary product of oxidative stress formed during lipid peroxidation and it is released as a result of toxic effect of reactive oxygen species in rats after BDL [26]. There are several reports indicating that levels of MDA increases after BDL in rats [19, 27, 28]. Results of the present study are consistent with previous works reporting high levels of MDA. In the present study, levels of MDA in the CAPE treated rats were significantly lower than in the BDL group. Although tissue MDA levels were clearly decreased by CAPE, its exact mechanism is not clear. A recent study reported by Kus *et al.* [29] showed that administration of CAPE decreased levels of MDA in rats exposed to carbon tetrachloride (CCl $_4$) induced hepatotoxicity. It is possible that the interference of CAPE with free radical

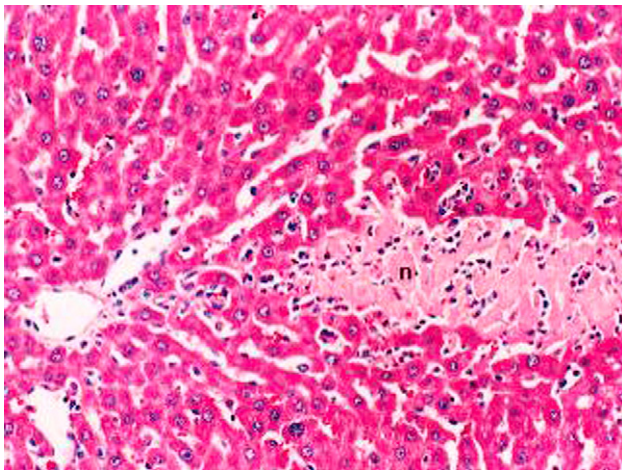


FIG. 1. BDL group. A large area of coagulation necrosis (n) is observed. H and E; $\times 20$. (Color version of figure is available online.)

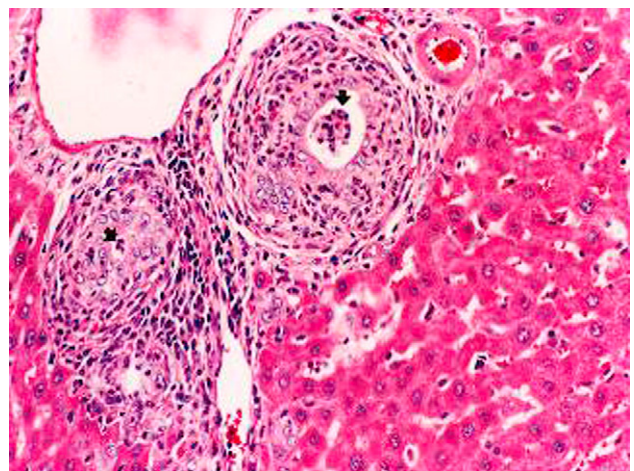


FIG. 3. BDL group. Periportal cell infiltration and cholangitis are observed. Bile ducts are filled with pus cells (arrows). H and E; $\times 20$. (Color version of figure is available online.)

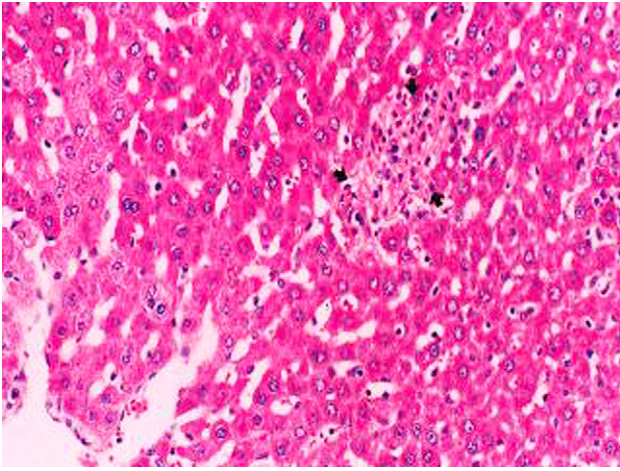


FIG. 4. BDL+CAPE group. A small necrotic hepatocyte group (arrows) between normal-appearing parenchymal cells are observed. H and E; $\times 20$. (Color version of figure is available online.)

generation is related to a decline oxidative stress in cholestatic rats.

GSH is an essential component of the cellular defense mechanism against oxidative stress that induced by ROS in rats with bile duct ligation [30]. GSH plays a pivotal role in the defense against oxidative stress, as a co-factor of glutathione peroxidases (selenium dependent and independent) participates in the elimination of hydrogen peroxide and lipid hydroperoxides [31]. In accordance with previous reports, our results showed depleted tissue GSH levels attributable to BDL [3, 32, 33]. Administration of CAPE restored GSH levels significantly. Restoration of tissue GSH levels by CAPE treatment could be partly related to its antioxidant and free-radical scavenging effect. Another explanation of this statistically significant increase in GSH levels seen in the CAPE treated rats compared with the BDL rats is the effect of CAPE upon the enzymes

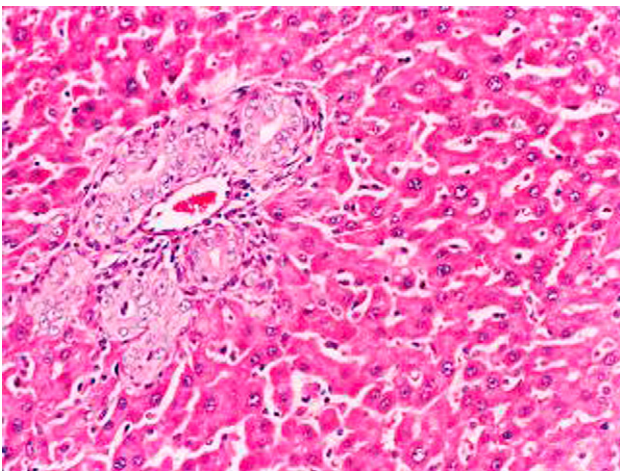


FIG. 5. BDL+CAPE group. Portal fibrosis is not evident. Bile duct proliferation is prominent. H and E; $\times 20$. (Color version of figure is available online.)

involved in GSH synthesis, whereby CAPE may maintain the levels of glutathione during oxidative stress [34]. This increased GSH is consistent with the protective effects of CAPE against oxidative damage in cholestasis, and that depletion of tissue GSH content enhances cellular injury caused by oxidative stress [35].

In the present study, we have observed many histopathological features of cholestasis such as focal necrosis, confluent necrosis, piecemeal necrosis, focal and portal inflammation, fibrosis, and apoptosis. HAI scores of the BDL group were significantly higher than that of the sham group. The degree of necro-inflammation showed significant difference between the BDL and CAPE treated groups. Portal fibrosis was not evident and histopathological evidence of parenchymatous injury was significantly reduced in the CAPE treated group.

In bile duct ligated rats; phospholipase is activated as a result of retention of bile salts, which are toxic for cells. Arachidonic acid products trigger ductal proliferation that is the initial phase of hepatic inflammation [36, 37]. As a result, several cytokines such as TNF- α from Kupffer cells and hepatocytes, IL-6, and IL-1 α are secreted [38, 39]. Both TNF- α and IL-6 are proinflammatory and have been studied specifically with respect to the pathophysiology of liver disease because many of their biological effects parallel the clinical and biochemical abnormalities found in patients with chronic liver disease [40]. The inhibitory effects of CAPE on IL-6 [41], IL-1 α , and TNF- α production [42] have been reported previously. In the current study, the levels of IL-6, IL-1 α , and TNF- α were significantly elevated due to BDL, while CAPE treatment markedly reduced these cytokine levels. Increased cytokine levels due to cholestasis, which tended to decrease with the treatment of CAPE, also supports the notion that CAPE ameliorates cholestatic liver injury through its antioxidant effect.

CONCLUSIONS

The present study demonstrates that intraperitoneal administration of CAPE maintains antioxidant defenses and reduces liver oxidative and cytokine damage, necro-inflammation (focal necrosis, confluent necrosis, piecemeal necrosis, focal and portal inflammation, and apoptosis) in bile ligated rats. This effect of Cape may be useful to preserve liver function in patients with biliary obstruction. However, more investigations are required to evaluate CAPE's antioxidant, anti-inflammatory, and hepatoprotective effect in clinical and experimental models.

REFERENCES

1. Sokol RJ, Devereaux M, Khandwala RA. Effect of dietary lipid and vitamin E on mitochondrial lipid peroxidation and hepatic injury in the bile duct-ligated rat. *Lipid Res* 1991;32:1349.

2. Hunt DR. The identification of risk factors and their application to the management of obstructive jaundice. *Aust NZ J Surg* 1980;50:476.
3. Gonzalez-Correa JA, De La Cruz JP, Martin-Aurioles E, et al. Effects of S-adenosyl-L-methionine on hepatic and renal oxidative stress in an experimental model of acute biliary obstruction in rats. *Hepatology* 1997;26:121.
4. Tomioka M, Inuma H, Okinaga K. Impaired Kupffer cell function and effect of immunotherapy in obstructive jaundice. *J Surg Res* 2000;92:276.
5. Parola M, Leonarduzzi G. On the role of lipid peroxidation in the pathogenesis of liver damage induced by long-standing cholestasis. *Free Radic Biol Med* 1996;20:351.
6. Sud'ina GF, Mirzoeva OK, Pushkareva MA, et al. Caffeic acid phenethyl ester as a lipoxygenase inhibitor with antioxidant properties. *FEBS Lett* 1993;329:21.
7. Hepsen IF, Bayramlar H, Gultek A, et al. Caffeic acid phenethyl ester to inhibit posterior capsule opacification in rabbits. *J Cataract Refract Surg* 1997;23:1572.
8. Ilhan A, Koltuksuz U, Ozen S, et al. The effects of caffeic acid phenethyl ester (CAPE) on spinal cord ischemia/reperfusion injury in rabbits. *Eur J Cardiothorac Surg* 1999;16:458.
9. Koltuksuz U, Ozen S, Uz E, et al. Caffeic acid phenethyl ester prevents intestinal reperfusion injury in rats. *J Pediatr Surg* 1999;34:34.
10. Ara C, Esrefoglu M, Polat A, et al. The effect of caffeic acid phenethyl ester on bacterial translocation and intestinal damage in cholestatic rats. *Dig Dis Sci* 2006;51:1754.
11. Pastor A, Collado PS, Almar M, et al. Microsomal function in biliary obstructed rats: Effects of S-adenosylmethionine. *J Hepatol* 1996;24:353.
12. Grunberger D, Banerjee R, Eisinger K, et al. Preferential cytotoxicity on tumor cells by caffeic acid phenethyl ester isolated from propolis. *Experientia* 1988;44:230.
13. Mihara M, Uchiyama M. Determination of malonaldehyde precursor in tissues by thiobarbituric acid test. *Anal. Biochem* 1978;86:271.
14. Wei H, Frenkel K. In vivo formation of oxidized DNA bases in tumor promoter-treated mouse skin. *Cancer Res* 1991;51:4443.
15. Gurlek A, Aydogan H, Parlakpınar H, et al. Protective effect of melatonin on random pattern skin flap necrosis in pinealectomized rat. *J Pineal Res* 2004;36:58.
16. Knodell RG, Ishak KG, Black WC, et al. Formulation and application of a numerical scoring system for assessing histological activity in asymptomatic chronic active hepatitis. *Hepatology* 1981;1:431.
17. Ishak K, Babbista A, Bianchi L, et al. Histological grading and staging of chronic hepatitis. *J Hepatol* 1995;22:696.
18. Krahenbuhl S, Talos C, Lauterburg BH, et al. Reduced antioxidative capacity in liver mitochondria from bile duct ligated rats. *Hepatology* 1995;22:607.
19. Canturk NZ, Canturk Z, Utkan NZ, et al. Cytoprotective effects of alpha tocopherol against liver injury induced by extrahepatic biliary obstruction. *East Afr Med J* 1998;75:77.
20. Ohta Y, Kongo M, Sasaki E, et al. Protective effect of melatonin against α -naphthylisothiocyanate-induced liver injury in rats. *J Pineal Res* 2000;29:15.
21. Levy R, Nagauker O, Sikuler E, et al. Elevated NADPH-oxidase activity in neutrophils from bile-duct-ligated rats: Changes in the kinetic parameters and in the oxidase cytosolic factor p47. *Biochim Biophys Acta* 1994;1220:261.
22. Tsuji K, Kubota Y, Yamamoto S, et al. Increased neutrophil chemotaxis in obstructive jaundice: An in vitro experiment in rats. *J Gastroenterol Hepatol* 1999;14:457.
23. Ohta Y, Kongo M, Kishikawa T. Melatonin exerts a therapeutic effect on cholestatic liver injury in rats with bile duct ligation. *J Pineal Res* 2003;34:119.
24. Ohta Y, Kongo M, Kishikawa T. Therapeutic effect of melatonin on cholestatic liver injury in rats with bile duct ligation. *Adv Exp Med Biol* 2003;527:559.
25. Gurel A, Armutcu F, Sahin S, et al. Protective role of alpha-tocopherol and caffeic acid phenethyl ester on ischemia-reperfusion injury via nitric oxide and myeloperoxidase in rat kidneys. *Clin Chim Acta* 2004;339:33.
26. Orellana M, Rodrigo R, Thielemann L, et al. Bile duct ligation and oxidative stress in the rat: Effects in liver and kidney. *Comp Biochem Physiol Toxicol Pharmacol* 2000;126:105.
27. Padillo FJ, Cruz A, Navarrete C, et al. Melatonin prevents oxidative stress and hepatocyte cell death induced by experimental cholestasis. *Free Radic Res* 2004;38:697.
28. Bulbuler N, Akkus MA, Cetinkaya Z, et al. Effects of melatonin and lactulose on the liver and kidneys in rats with obstructive jaundice. *Pediatr Surg Int* 2002;18:677. Epub 2002 Oct 26.
29. Kus I, Colakoglu N, Pekmez H, et al. Protective effects of caffeic acid phenethyl ester (CAPE) on carbon tetrachloride-induced hepatotoxicity in rats. *Acta Histochem* 2004;106:289.
30. Peres W, Tunon MJ, Collado PS, et al. The flavonoid quercetin ameliorates liver damage in rats with biliary obstruction. *J Hepatol* 2000;33:742.
31. Spitz DR, Sullivan SJ, Malcolm RR, et al. Glutathione dependent metabolism and detoxification of 4-hydroxy-2-nonenal. *Free Radic Biol Med* 1991;11:415.
32. Ebrahimkhani MR, Sadeghipour H, Dehghani M, et al. Homocysteine alterations in experimental cholestasis and its subsequent cirrhosis. *Life Sci* 2005;76:2497.
33. Cruz A, Padillo FJ, Granados J, et al. Effect of melatonin on cholestatic oxidative stress under constant light exposure. *P. Cell Biochem Funct* 2003;21:377.
34. Ihan A, Akyol O, Gurel A, et al. Protective effects of caffeic acid phenethyl ester against experimental allergic encephalomyelitis-induced oxidative stress in rats. *Free Radic Biol Med* 2004;37:386.
35. Meister A, Anderson ME. Glutathione. *Annu Rev Biochem* 1983;52:711.
36. Reilly PM, Schiller HJ, Bulkley GB. Pharmacologic approach to tissue injury mediated by free radicals and other reactive oxygen metabolites. *Am J Surg* 1991;161:488.
37. Muriel P, Suarez R. Role of lipid peroxidation in biliary obstruction in the rat. *J Appl Toxicol* 1994;14:423.
38. Bemelmans M, Gouma DJ, Greve JW, et al. Cytokines tumor necrosis factor and interleukin-6 in experimental biliary obstruction in mice. *Hepatology* 1992;15:1132.
39. Sewnath ME, Van Der Poll T, Ten Kate FJ, et al. Interleukin-1 receptor type I gene-deficient bile duct-ligated mice are partially protected against endotoxin. *Hepatology* 2002;35:149.
40. Lee FY, Lu RH, Tsai YT, et al. Plasma interleukin-6 levels in patients with cirrhosis. Relationship to endotoxemia, tumor necrosis factor-alpha, and hyperdynamic circulation. *Scand J Gastroenterol* 1996;3:500.
41. Xiao W, Hodge DR, Wang L, et al. Cooperative functions between nuclear factors NF- κ B and CCAT/enhancer-binding protein- β (C/EBP- β) regulate the IL-6 promoter in autocrine human prostate cancer cells. *Prostate* 2004;61:354.
42. Natarajan K, Singh S, Burke TR, Jr., et al. Caffeic acid phenethyl ester is a potent and specific inhibitor of activation of nuclear transcription factor NF- κ B. *Proc. Nat. Acad. Sci. USA* 1996;93:9090.